

## Technical Data Sheet

**Codex ACTOne™ Non-Wash Calcium Dye Kit**

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**Product Information**

**Catalog Number:** CB-80500-305  
**Size:** Reagents for 50 plates  
**Components:** Codex Calcium Dye, 5 vials, lyophilized (Part No: 80500-113)  
10X Calcium Dye Signal Enhancer, 50 ml (Part No: 80500-114)

**Description**

The Codex ACTOne™ Non-Wash Calcium Dye Kit allows homogeneous measurement of intracellular calcium changes caused by activation of G-protein coupled receptors or calcium channels. The assay involves only one step of dye addition and does not require any washing steps. It is user friendly and cost effective. The assay can be easily implemented in a high throughput environment.

**Storage**

Codex Calcium Dye	-20°C (protected from light)
10X Calcium Dye Signal Enhancer	Room Temp.

**Materials not included**

DMSO	Sigma D4540
Probenecid	Sigma P8761

## ASSAY PROTOCOL

### Prepare the cell plate:

1. Seed 80  $\mu$ l of cell suspension into each well of a 96-well plate or 20  $\mu$ l of cell suspension into each well of a 384-well plate.
2. Grow the cells overnight in a CO<sub>2</sub> incubator

### Prepare the buffers:

On the 2<sup>nd</sup> day:

1. Prepare Buffer A (1X HBSS with 20 mM HEPES):  
10 ml of 1M HEPES, pH 7.3 + 490 ml of 1X HBSS
2. Prepare 1 ml of 500 mM Probenecid.  
Dissolve 142 mg of Probenecid in 1 ml of 1N NaOH
3. Prepare stock solution of calcium dye  
Add 80  $\mu$ l of DMSO into each well containing 0.5 mg of calcium dye
4. Prepare **2X Dye Loading Buffer** (1 plates).  
Add 8 ml of Codex 10X Calcium Dye Signal Enhancer into 72 ml of Buffer A.  
Add 800  $\mu$ l of 500 mM Probenecid.  
Add 80  $\mu$ l of calcium dye stock solution. Mix well by vortexing.

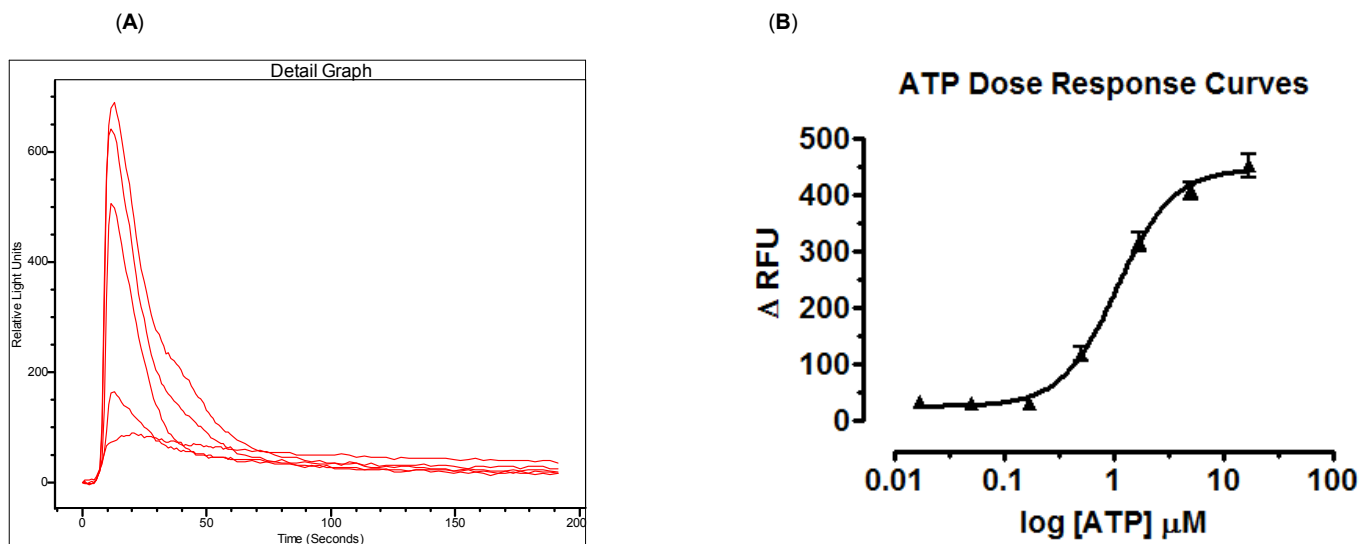
### Assay:

1. Take the cell plate out from the incubator.
2. Add same volume of **2X Dye Loading Buffer** into each well, 80  $\mu$ l to a 96-well plate or 20  $\mu$ l to a 384-well plate.
3. Incubate at 37 °C incubator for 1 hr.
4. Take the cells out of the incubator and leave at room temp (in the dark) for 30 min.
5. Put the plate into the instrument for assay

For assays performed on a FlexStation (MDS), use the following wavelength parameters. Excitation: 485 nm; Emission: 530 nm; Cutoff 515 nm

**Note.** *Dispense speed and height for compound additions need to be optimized for each instrument.*

## Appendix



**Figure 1. Response of endogenous P2Y receptors to ATP.** HEK293 cells were plated overnight in 20  $\mu$ l culture medium on a 384 well Biocoat poly-D lysine coated plate. The next day, the cells were dye-loaded by adding 20  $\mu$ l of 2X Dye Loading Buffer and incubating for 1 hour at 37°C. ATP solution was added (10  $\mu$ l/well) by a FLIPR Tetra (Molecular Devices), and the data was recorded simultaneously. **A.** Kinetic curve of calcium response to different concentrations of ATP. **B.** ATP dose response curve (n = 4). EC50 = 1.1  $\mu$ M.

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